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Bio-Polyurethane Resins Derived from Liquid Fractions of Lignin for the Modification of Ramie Fibers

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ABSTRACT

In this study, technical lignin from black liquor was used as a pre-polymer for the preparation of bio-polyurethane (Bio-PU) resins. Briefly, the isolated lignin was fractionated using ethyl acetate (EtAc) and methanol (MeOH). The liquid fractions of lignin, such as lignin-EtAc (L-EtAc) and lignin-methanol (L-MeOH), were mixed with 10% of polymeric isocyanate (based on the weight of liquid fractions) to obtain Bio-PU resins. The isolated lignin, fractionated lignin, and lignin-derived Bio-PU resins were characterized using several techniques. The obtained Bio-PU resins were then used to modify ramie fibers using vacuum impregnation method. Fourier Transform Infrared (FTIR) spectroscopy, Differential Scanning Calorimetry (DSC), and Thermogravimetric Analysis (TGA) revealed that the isolated lignin had quite similar characteristics to the lignin standard. Fractionation of lignin with EtAc and MeOH altered its characteristics. FTIR, DSC, and TGA showed that solid fractions of lignin had similar characteristics to lignin standard and isolated lignin, while the liquid fractions had characteristics from lignin and the solvents. The absorption band of isocyanate ($-N=C=O$) groups was shifted to 2285 cm^{-1} from 2240 cm^{-1} owing to the reaction with the $-OH$ groups in lignin, forming urethane ($R-NH-C=O-R$) groups at 1605 cm^{-1} in Bio-PU resins. Thermal properties of Bio-PU resins derived from L-EtAc exhibited greater endothermic reaction compared to Bio-PU-L-MeOH. As a result, the free $-N=C=O$ groups in Bio-PU resins have reacted with $-OH$ groups on the surface of ramie fibers and improved its thermal properties. Modification of ramie fibers with Bio-PU resins improved the fibers' thermal stability by 15% using Bio-PU-LEtAc for 60 min of impregnation.

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1. Introduction

Lignin is the second most abundant biopolymer after cellulose obtained from agroforestry biomass. It is found in all lignocellulosic plants and is derived after deposition of the carbohydrates (Boerjan et al. 2003). Lignin is regarded as a key raw material to achieve a sustainable forest product bioeconomy, particularly for the pulp and paper industry (Wenger et al. 2020). Lignin is a renewable, amorphous, and aromatic biopolymer, isolated from black liquor which is a by-

product of the pulp and paper industry (Krutov et al. 2014). The pulp and paper industry worldwide produces an estimated 40–50 million tons of lignin annually (FAO 2019). The chemical composition of black liquor depends on the source of the raw fiber and therefore determines its physical properties. The high solids content and availability of black liquor as the main by-product of the pulp and paper industry make great use of black liquor as a strategic and promising raw material for lignin (Antov et al. 2021). This isolated lignin from black liquor is referred to technical lignin (El Mansouri and Salvadó 2007). Lignin provides greater prospects for higher added value applications in renewable products. Lignin polymers contain several important functional groups, including phenolic hydroxyl, aliphatic hydroxyl, and carboxylic acid groups (Antov et al. 2020a; Solihat et al. 2021). The reactivity of different lignins depends on their chemical structure. Therefore different lignin samples contain different types and numbers of these functional groups, which can be used to manufacture industrial biomaterials and biocomposites (Antov et al. 2020b; Falah et al. 2020).

Technical lignin can be converted to the industrial one by fractionation. Technical lignin has broad molecular weight distribution, between 1000 g/mole and 100,000 g/mole, makes it difficult to commercially utilize lignin (Toledano et al. 2010). Fractionation of lignin has two potential benefits: chemical structure versus bulk property relationship can be obtained, and the fractions themselves can have properties useful for various applications (Velez and Thies 2013). There are three available fractionation methods of lignin. The first one is differential precipitation, which extracts lignin from black liquor by decreasing the pH gradually, using a strong mineral acid (García et al. 2009). This method produces a high yield of lignin, but due to the formation of colloids during precipitation, it results in relatively low purity of the isolated lignin. The second method uses selective solvents for the lignin extraction, with the help of enzymes, catalysts, and ionic liquids to obtain low molecular weight lignin (Toledano et al. 2010). This method allows to produce high added value products from the isolated lignin but with very high production costs. The last method utilizes membrane technology to obtain lignin fractions with specific molecular weight distributions without using chemicals (Jönsson et al. 2008). The fractions of lignin are more suitable to be used as building blocks for bio-based polymers such as bio-based polyurethane (PU) resins (Griffini et al. 2015; Wang et al. 2019). Cross-linked PU resins polymer from the liquid fractions of lignin has good thermal and thermomechanical properties and a good shape memory effect. PU resins polymer is one of the most useful and multi-purpose polymers with formulation covering a wide range of products and applications. So far, many types of polyols and diisocyanates have been explored to prepare the PU resins. Compared with petroleum polyol-based materials, lignin-based polyols are more environmentally friendly, biodegradable, cost less, and exhibit potential application for developing economic and sustainable PU resins (Alinejad et al. 2019).

Ramie (*Boehmeria nivea* (L.) Gaudich) fibers are bast fibers, which are extensively available in nature, native in China, and widespread in Asia (Angelini et al. 2000). Ramie fibers are known as one of the fully biodegradable natural fibers, characterized by a high strength (61–128 GPa), small diameter (10–60 μm), low specific density (1.5 g/cm^3), antibacterial properties, higher absorbency, air permeability, and lower wrinkle characteristics, with higher cellulose content of approximately 65–75% compared to other bast fibers (Angelini et al. 2015; Du et al. 2015; Li et al. 2015; Yuan et al. 2016). Ramie fibers have been used as raw material for composites and textiles to replace synthetic fiber-oriented composites due to their outstanding biodegradability, renewability, decomposability, stiffness, higher length-to-weight ratio, and low cost (Angelini et

al. 2015; Djafar et al. 2020). However, the negative characteristics of ramie fibers are flammability and thermal instability (Dorez et al. 2013).

This research is aimed to prepare and characterize bio-polyurethane (Bio-PU) resins from liquid fractions of lignin and use the Bio-PU resins for modification of ramie fibers. In this study, the black liquor obtained from the pulp and paper industry was used as starting material for isolated lignin. The isolated lignin was fractionated prior to the preparation of Bio-PU resins. Ramie fibers were modified using the obtained Bio-PU resins. Several techniques were employed to characterize the isolated lignin, fractionated lignin, Bio-PU resins, and ramie fibers.

2. Materials and Methods

2.1. Materials

Black liquor of *Acacia mangium* kraft pulping with 70% solids content was obtained from PT. Tanjung Enim Lestari Pulp and Paper (PT. TELPP, South Sumatera, Indonesia). For comparison, commercial lignin from Sigma Aldrich was also used to a similar design condition. HCl (37% w/v) was used for lignin isolation process. Organic solvents, namely ethyl acetate (EtAc) and methanol (MeOH), were used for lignin fractionation. Standardized kraft lignin from Sigma Aldrich (CAS No. 8068-05-1) was used as a standard. Polymeric 4,4-methane diphenyl diisocyanate (pMDI, 31% NCO content) was purchased from Anugrah Raya Kencana Company (ARK, Banten, Indonesia) for the preparation of bio-polyurethane pre-polymer. Degummed ramie fibers were obtained from a local farmer at CV. RABERSA (Wonosobo, Central Java, Indonesia).

2.2. Methods

2.2.1. Isolation and fractionation of lignin

Lignin was isolated by single step precipitation. Approximately 1M of HCl was added to 800 g of the black liquor, which has a pH of about 12, up to the solution reached pH of 2. Lignin suspensions in the acid solution obtained by the previous method were kept for 24 h to complete the precipitation process. Afterward, their solution was separated from the lignin precipitates by pipette, washed three times with distilled water, and then put in the freezer for 24 h. After that, the lignin precipitates were separated from their solution by vacuum filtration and Whatman filter paper no 4, then dried in the oven at 45°C or 24 h. The yield and moisture content of lignin were characterized according to the published work (Hermiati et al. 2017).

Liquid fractions of lignin were obtained by fractionation method using two different solvents, i.e., ethyl acetate and methanol. Lignin was mixed with ethyl acetate (EtAc) with an EtAc:Lignin ratio of 15:1 in a beaker. The fractionation process was carried out for 24 h at room temperature with a stirring speed of 200 rpm. After the fractionation process was complete, the insoluble lignin solid EtAc (solid residue) was separated with EtAc soluble lignin (liquid fraction) by vacuum filtration and Whatman filter paper no. 4. Similar methods were also carried out for lignin fractionation with methanol (L-MeOH) solvents. Those liquid fractions of lignin were used as bio-polyurethane resins.

2.2.2. Preparation of bio-polyurethane resins

Two types of bio-polyurethane (Bio-PU) resins were produced using the liquid fractions of lignin, namely L-EtAc and L-MeOH. pMDI was diluted with acetone until reaching 50% of

concentration and then was used as a component for bio-polyurethane pre-polymer. Approximately 25 mL of each liquid fraction of lignin was mixed with 10% of diluted pMDI based on the weight of the liquid fractions in 100 mL of beaker glass under 300 rpm of stirring speed. The reaction was carried out under rapid stirring of 500 rpm at $25^{\circ}\text{C} \pm 2^{\circ}\text{C}$ for 1 h, as shown in **Fig. 1**.

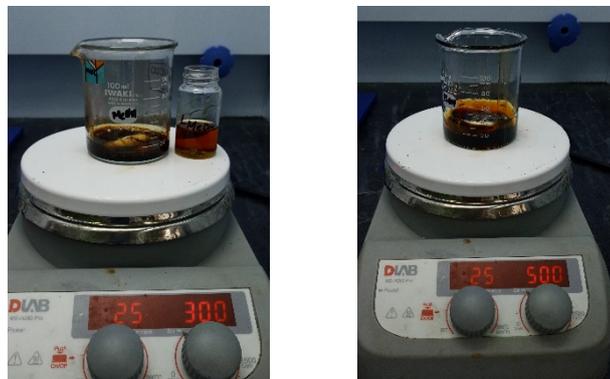


Fig. 1. Preparation of Bio-PU resins derived from liquid fractions of lignin.

2.2.3. Characterization of lignin and bio-polyurethane resins

Functional groups of isolated lignin, fractionated lignin, and Bio-PU resins were investigated using Fourier Transform Infra-Red (FTIR) spectroscopy (SpectrumTwo, Perkin Elmer Inc., USA) by the Universal Attenuated Total Reflectance (UATR) method. Lignin and its fractions samples were scanned from $4000\text{--}400\text{ cm}^{-1}$ at $23^{\circ}\text{C} \pm 2^{\circ}\text{C}$ with an accumulative of 16 scans. Each spectrum was normalized using min-max normalization in Spectrum software (Ver. 10.5.3, Perkin Elmer Inc., USA).

Thermal behavior of isolated lignin, fractionated lignin, and Bio-PU resins were analyzed using Differential Scanning Calorimetry (DSC 4000, Perkin Elmer Inc., USA). Around $5\text{ mg} \pm 0.5\text{ mg}$ of each sample was put in a standard aluminum pan and heated from 0°C to 300°C at a heating rate of $10^{\circ}\text{C}/\text{min}$ under nitrogen flow of $20\text{ mL}/\text{min}$. The endothermic peak temperature (T_p) of each sample was detected using Pyris software (Ver. 13.3.1, Perkin Elmer Inc., USA).

Thermal stability of isolated lignin, fractionated lignin, and Bio-PU resins were investigated using Thermogravimetric Analyzer (TGA 8000, Perkin Elmer Inc., USA). Each sample was heated from 25°C to 750°C at a heating rate of $10^{\circ}\text{C}/\text{min}$. Nitrogen gas was purged with a flow of $20\text{ mL}/\text{min}$ during the analysis. Weight loss and its derivative thermogravimetric (DTG) were calculated using Pyris software (Ver. 11.1.1, Perkin Elmer Inc., USA).

2.2.4. Impregnation of Ramie Fibers With Bio-Polyurethane Resins

Degummed ramie fibers were impregnated with Bio-PU-L-EtAc and Bio-PU-L-MeOH resins. Impregnation was performed in 1L vacuum chambers with a 2-stage vacuum pump (VC0918SS, VacuumChambers.us., USA) as depicted in **Fig. 2**. The initial weight of ramie fibers was recorded prior to vacuum impregnation. Approximately 5 g of ramie fibers were immersed in 50 mL of Bio-PU resins and vacuum impregnated at $25^{\circ}\text{C} \pm 2^{\circ}\text{C}$ under 50 kPa for 30 and 60 min. Then, the impregnated fibers were dried in an oven at 60°C for 24 h. The dried ramie fibers were weighed to determine the weight gain after impregnation. The weight gain (%) was calculated by

dividing the mass of ramie fibers after impregnation with the initial mass of ramie fibers. The impregnated ramie fibers were then stored in zip-lock plastic bags for further examination.



Fig. 2. A set of vacuum chamber for impregnation of ramie fibers with Bio-PU resins.

2.2.5. Evaluation of ramie fibers properties

Functional groups of ramie fibers before and after impregnation with Bio-PU resins were investigated using Fourier Transform Infra-Red (FTIR) spectroscopy (SpectrumTwo, Perkin Elmer Inc., USA) with Universal Attenuated Total Reflectance (UATR) method. The fibers were scanned from $4000\text{--}400\text{ cm}^{-1}$ at $23^\circ\text{C} \pm 2^\circ\text{C}$ with an accumulative of 16 scans. Each spectrum was normalized using min-max normalization in Spectrum software (Ver. 10.5.3, Perkin Elmer Inc., USA).

Thermal behavior of ramie fibers before and after impregnation with Bio-PU resins were analyzed using Differential Scanning Calorimetry (DSC 4000, Perkin Elmer Inc., USA). Around $5\text{ mg} \pm 0.5\text{ mg}$ of each sample was put in a standard aluminum pan and heated from 0°C to 300°C at a heating rate of $10^\circ\text{C}/\text{min}$ under nitrogen flow of $20\text{ mL}/\text{min}$. The endothermic and exothermic peak temperature (T_p) of each sample was detected using Pyris software (Ver. 13.3.1, Perkin Elmer Inc., USA).

Thermal stability of ramie fibers before and after impregnation with Bio-PU resins was investigated using Thermogravimetric Analyzer (TGA 8000, Perkin Elmer Inc., USA). Each sample was heated from 25°C to 750°C at a heating rate of $10^\circ\text{C}/\text{min}$. Nitrogen gas was purged with a flow of $20\text{ mL}/\text{min}$ during the analysis. Weight loss and its derivative thermogravimetric (DTG) were calculated using Pyris software (Ver. 11.1.1, Perkin Elmer Inc., USA).

3. Results and Discussion

3.1. Characteristics of Lignin

FTIR spectroscopy of lignin standard and isolated lignin are shown in **Fig. 3**. Lignin standard and isolated lignin exhibited a broad absorption band at 3365 cm^{-1} that represented the O-H stretching vibrations in phenolic and aliphatic O-H groups, $2930\text{--}2825\text{ cm}^{-1}$ assigned to C-H stretching of CH_3 and CH_2 , 1705 cm^{-1} attributed to C=O stretching of unconjugated ketone, carbonyl, and ester groups, and $1595\text{--}1510\text{ cm}^{-1}$ represented to C-C stretching of aromatic ring (Jung et al. 2015; Tejado et al. 2007). The main difference between lignin standard and isolated lignin was at wavenumber of 1080 cm^{-1} , which was only detected in the lignin standard. This band was attributed to C-O groups of primary alcohol, such as $-\text{CH}_2\text{OH}$, which was probably decomposed during acid precipitation of isolated lignin from black liquor. This study showed that

lignin isolated from black liquor was similar to the lignin standard in terms of functional groups. The average yield of isolated lignin was around 28.8%, showing that approximately 28.8 g of lignin can be obtained from 100 g of black liquor. The average moisture content of lignin was around 5.9%, which is two times higher than that of the lignin standard. These results are in agreement with previously published works (Jung et al. 2015; Tejado et al. 2007).

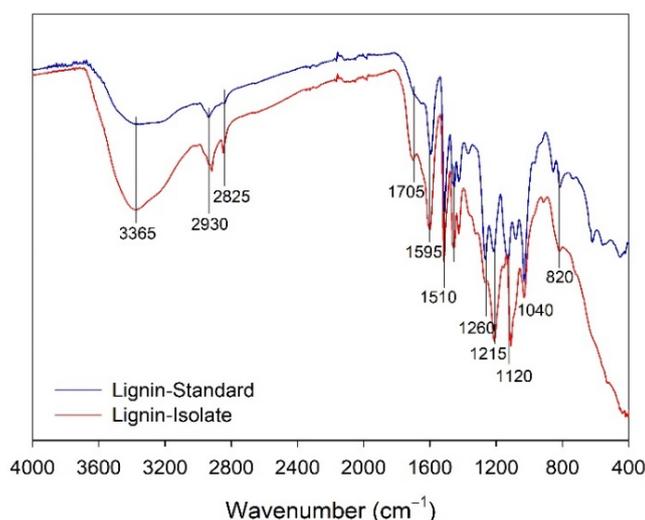


Fig. 3. Typical FTIR spectra of lignin.

Fractionation of lignin with EtAc resulted in several new functional groups (**Fig. 4a**). EtAc had typical functional groups at bands 3365 cm^{-1} with a low intensity that representing O–H stretching, low absorption band for C–H stretching was exhibited in the area below 3000 cm^{-1} . Sharp absorption band for C=O stretching is shown at 1740 cm^{-1} and 1375 cm^{-1} representing C–H bending (CH_3), while the absorption band for CO ethanoate appears at 1230 cm^{-1} . A strong absorption band for CO–O–CO stretching is shown at 1040 cm^{-1} . FTIR spectra further reveals that solid fraction of lignin after fractionation with ethyl acetate (L-EtAc) had similar functional groups with lignin standard and EtAc. This indicated that the fractionation process had been successfully carried out. It is identified by the presence of functional groups whose intensity is higher than standard lignin and solvents (EtAc) used at bands 3365 , 2930 , and 2825 cm^{-1} . Besides that, there are also absorption bands at 1705 cm^{-1} , 1595 cm^{-1} , 1510 cm^{-1} , 1230 cm^{-1} , and 1040 cm^{-1} . While the liquid fraction (L-EtAc-liquid) had absorption bands that identify the functional groups of lignin and EtAc. Some of the absorption bands have shifted in wavenumber, e.g., 3650 cm^{-1} (O–H stretching) and 2985 cm^{-1} . There are more absorption bands around $3000\text{--}2800\text{ cm}^{-1}$, showing that many methyl and methylene groups are fragmented during the fractionation process. A sharp absorption band at 1740 cm^{-1} representing C=O stretching, 1375 cm^{-1} to C–H bending, 1230 cm^{-1} to C=O stretching, and 1040 cm^{-1} to CO–O–CO stretching (anhydride), but in this spectra, there are several absorption bands of lignin missing because it is closed by liquid, for example in the 1595 cm^{-1} and 1510 cm^{-1} that representing C–C stretching vibration. These results were in agreement with the published works (Chauhan et al. 2014; Griffini et al. 2015).

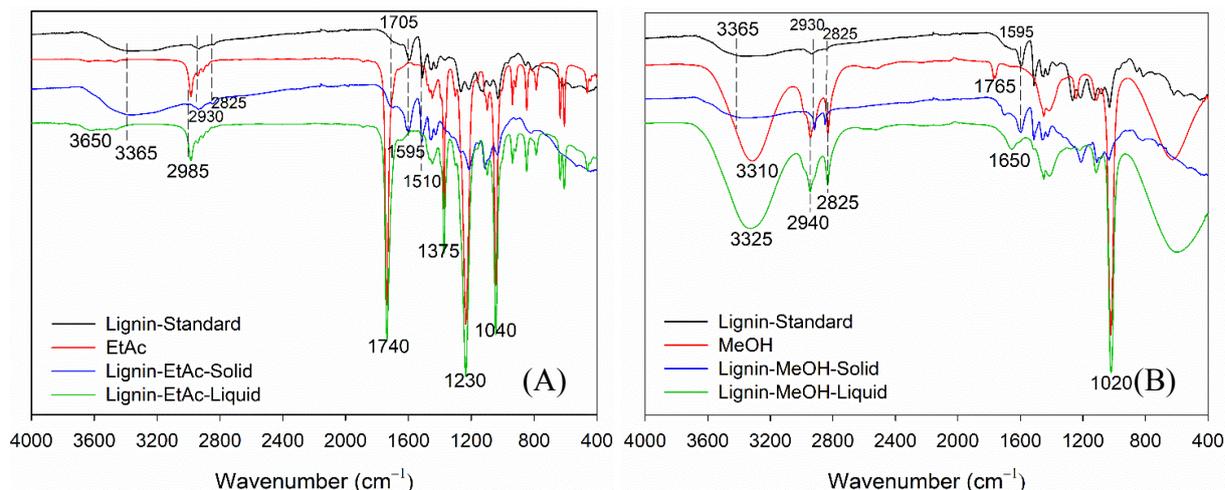


Fig. 4. Comparison of FTIR spectra of isolated and fractionated lignin: (a) fractionated with ethyl acetate (EtAc) and (b) fractionated with methanol (MeOH).

Fig. 4b presents FTIR spectra and absorption band assignments of fractionated lignin with methanol (MeOH). In general, MeOH has a medium intensity absorption band at 3310 cm^{-1} assigned to the O–H stretching (alcohol), 2940 and 2825 cm^{-1} are attributed to the C–H stretching, also in the fingerprint area, it has a sharp absorption band at 1020 cm^{-1} that represents C–O stretching. Furthermore, there is a small absorption peak at 1765 cm^{-1} that represents C=O stretching of pure MeOH. However, the peak is no longer visible in the spectra L–MeOH-solid or L–MeOH-liquid. The fractionation process lignin with ethanol is seen in the spectra L–MeOH-solid and L–MeOH-liquid that is identified by the presence of functional groups around at $3550\text{--}3200\text{ cm}^{-1}$, $3000\text{--}2840\text{ cm}^{-1}$, $2830\text{--}2695\text{ cm}^{-1}$, and $1650\text{--}1566\text{ cm}^{-1}$. In these spectra, there are absorption bands between $3000\text{--}2695\text{ cm}^{-1}$ are attributed to the C–H stretching vibrations of the methyl and methylene groups. It is identified that many methyl groups are fragmented than before (lignin without fractionation process), but there are absorption bands in the fingerprint area that are not visible in L–MeOH-solid and appear at the L–MeOH-liquid, e.g., on the absorption band at 1020 cm^{-1} . These results were in agreement with the published works (Chauhan et al. 2014; Griffini et al. 2015).

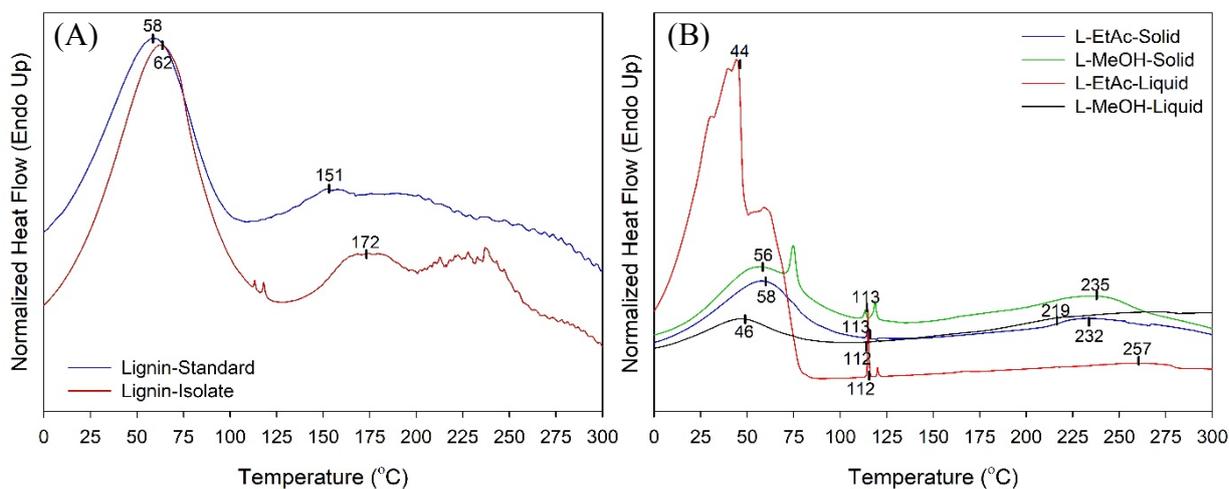


Fig. 5. DSC thermograms of lignin: (a) before fractionation and (b) after fractionation.

The thermal behavior of lignin was investigated using DSC analysis. In general, lignin had two endothermic reactions detected in DSC thermograms (**Fig. 5a**). The first endothermic peak (T_{p1}) occurred at around 58°C for lignin standard and 62°C for isolated lignin. During the first endothermic reaction, lignin absorbed heat from the environment to evaporate moisture. The isolated lignin had a bit higher T_{p1} due to its moisture content that was greater than the lignin standard. The second endothermic reaction was detected at T_{p2} of 151°C for lignin standard and 172°C for isolated lignin. At this temperature, lignin has been plasticized, which deformed the lignin and reduced its rigidity. This phenomenon is also known for glass transition (T_g), where lignin generally has greater T_g than other synthetic polymers at around 100°C to 180°C (Buranov et al. 2010). The high T_g of lignin is because of hydrogen bonds between –OH of phenolic groups of lignin. Several studies reported that commercial kraft lignin has T_g of around 93°C to 120°C, softwood's lignin has an average T_g of 158°C, and hardwood's lignin has an average T_g of 161°C (Cui et al. 2013; Jung et al. 2015). The degradation of lignin started from a temperature of 200°C.

By contrast, fractionated lignin showed different thermal behaviors with the lignin standard and isolated lignin (**Fig. 5b**). The effect of solvent on lignin fractions occurred from the beginning of the endothermic reaction, where EtAc and MeOH volatilized at low temperature of 44°C to 46°C. As the results, the T_{p2} of lignin fractions, which corresponded to T_g , was lower compared to the lignin standard and isolated lignin. T_{p2} was detected at the temperature of around 112°C to 113°C. Lower T_g of lignin fractions was obviously due to depolymerization of lignin structure. Fractionation broken down lignin macromolecules into smaller molecules. Degradation of lignin fractions was detected around 219°C to 257°C. These results were in agreement with the published works (Chauhan et al. 2014; Griffini et al. 2015).

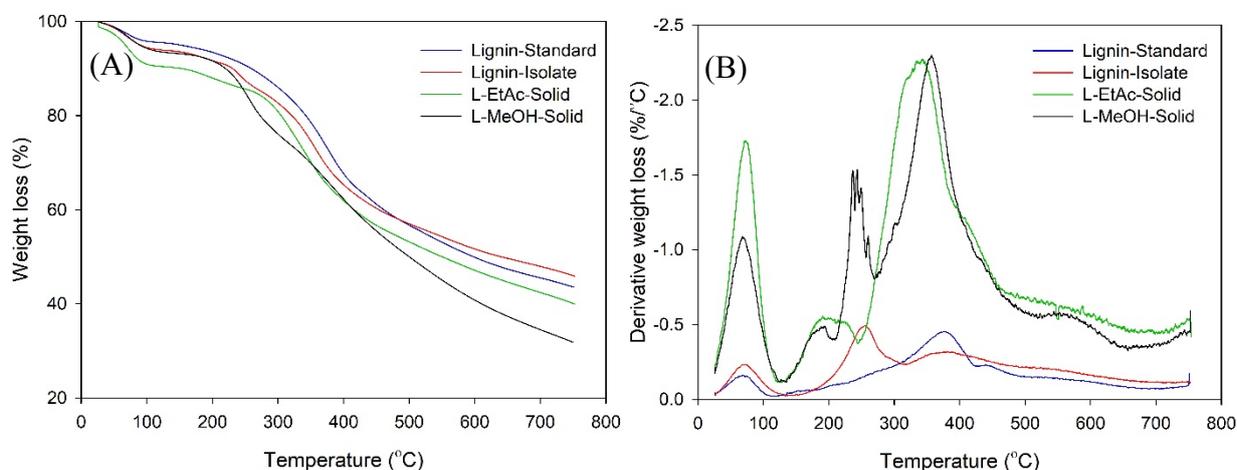


Fig. 6. Thermal stability of lignin: (a) TGA, (b) DTG.

The thermal stability of lignin was monitored by means of TGA-DTG (**Fig. 6**). TGA-DTG thermograms showed that three thermal degradations were detected during the analysis. The first degradation occurred at 65°C for lignin standard and at 70°C for isolated lignin owing to the evaporation of moisture in lignin. This resulted in around 5-7% of weight loss from the lignin sample. This result was supported by a DSC analysis, which showed an endothermic reaction, occurred at around 58°C to 62°C (**Fig. 6**). The second degradation was detected at a temperature of 120°C to 480°C, which was assigned to the lignin-carbohydrates' degradation. Initial degradation occurred at 120°C to 260°C mostly due to decomposition of carbohydrates, and then followed by degradation of lignin starting from 200°C to 480°C (Mimini et al. 2019). At this stage,

around 32-35% of the weight of the lignin sample was lost due to thermal degradation. The last degradation was observed above 500°C, attributed to the decomposition of the aromatic ring structure of lignin. The lignin sample was further heated to 750°C, leaving residue of around 43% to 46% from the initial weight. This indicates that non-volatile compounds in lignin are converted to condensed aromatic structure and become residue at 750°C (Tejado et al. 2007).

3.2. Properties of Bio-polyurethane Resins

Functional groups of Bio-PU resins and their raw materials are displayed in **Fig. 7a**. As mentioned in the previous section, the L-EtAc-liquid fraction had an absorption band at 2985 cm⁻¹ that corresponded to C–H stretching, while pMDI had the sharp peak at 2240 cm⁻¹ associated with isocyanate groups (–N=C=O) stretching. The reaction between L-EtAc-liquid and pMDI resulted in Bio-PU resins with specific functional groups. The spectrum of Bio-PU resins obtained from L-EtAc-liquid with pMDI showed CH₃ and CH₂ stretching that identified at 2985 cm⁻¹. In addition, the absorption band of free –N=C=O groups was shifted to 2285 cm⁻¹ from 2240 cm⁻¹. The shifting of –N=C=O groups, due to the reaction with the –OH groups in L-EtAc-liquid, formed urethane groups (R–NH–C=O–R) at a weak absorption band at 1605 cm⁻¹ (Lubis et al. 2019, 2020). However, the presence of free –N=C=O groups at 2285 cm⁻¹ indicated that not all pMDI has reacted with L-EtAc-liquid fraction to form Bio-PU resins. The results were in agreement with published work that used technical grade of kraft lignin to prepare PU resins (Nacas et al. 2017).

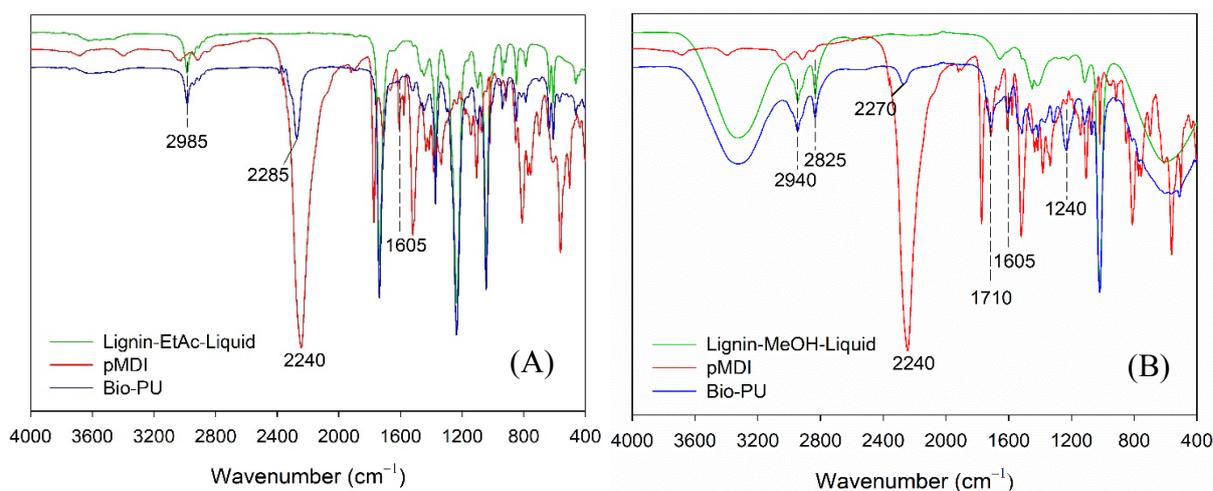


Fig. 7. FTIR spectra of liquid bio-polyurethane (Bio-PU) resins derived from fractionated lignin: (a) fractionated with ethyl acetate (EtAc), (b) fractionated with methanol (MeOH)

As depicted in **Fig. 7b**, Bio-PU resins have absorption bands at 2940 cm⁻¹ and 2825 cm⁻¹, corresponds to C–H stretching (CH₃ and CH₂). These absorption bands were also observed in L-MeOH-liquid. In addition, Bio-PU resins also had free –N=C=O groups at wavenumber of 2270 cm⁻¹, shifting from 2240 cm⁻¹. The peak of free –N=C=O groups in Bio-PU-L-MeOH was smaller than that in Bio-PU-L-EtAc. This indicates that pMDI has reacted more with –OH groups in L-MeOH-liquid to form urethane groups (R–NH–C=O–R) at 1605 cm⁻¹ than that of L-EtAc-liquid. The band at 1710 cm⁻¹ was assigned to C=O of urethane in Bio-PU resins. The wavenumber of 1240 cm⁻¹ was associated with C–N stretching of urethane in Bio-PU resins. This result was in agreement with the published work (Nacas et al. 2017). Based on the result, the fractionation

process was successfully used to prepare Bio-PU resins from fractionated lignin by reacting it with pMDI, where L-MeOH-liquid was more suitable as the Bio-PU pre-polymer.

Thermal properties of Bio-PU resins were monitored using DSC and TGA-DTG analysis. DSC thermograms of Bio-PU resins showed a T_g at around 133°C and an endothermic peak at T_p of 272°C for Bio-PU-L-EtAc, and a T_g at 142°C and an endothermic peak at T_p of 274°C for Bio-PU-L-MeOH (Fig. 8a). The T_g of Bio-PU-L-EtAc was lower than that of Bio-PU-L-MeOH, indicating that the Bio-PU-L-MeOH had a more hard segment due to the urethane formation. Hard segment in PU resins is originated from urethane groups from the reaction of pMDI and polyol (Somdee et al. 2019). A similar trend was observed for the endothermic peak, which was attributed to the melting of Bio-PU resins. Bio-PU-L-EtAc had a lower T_p of melting than that of Bio-PU-L-MeOH. This could be due to that Bio-PU-L-MeOH had more urethane groups and eventually produced more hard segments compared to Bio-PU-L-EtAc.

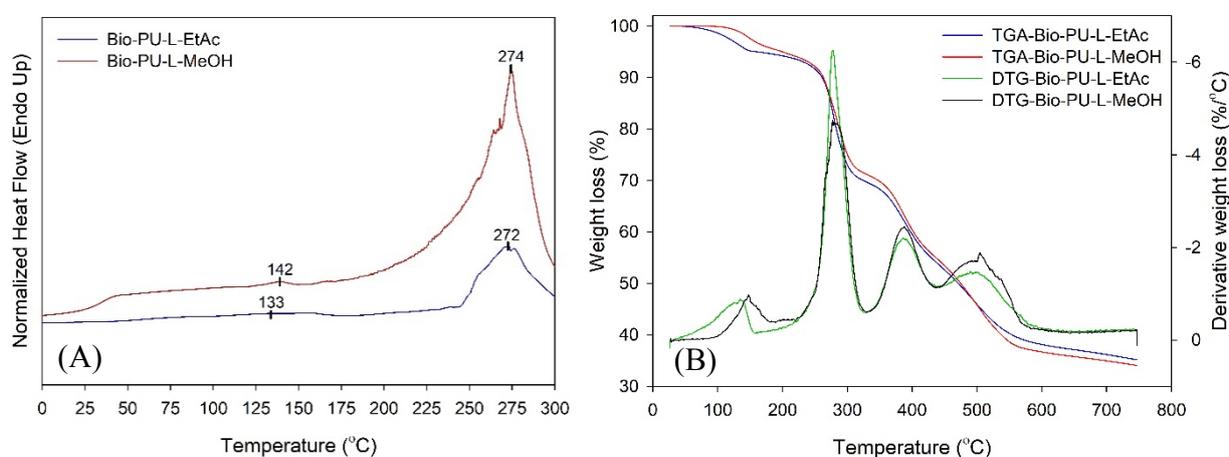


Fig. 8. Thermal properties of Bio-PU derived from lignin: (a) DSC, (b) TGA-DTG.

TGA-DTG analysis revealed that Bio-PU resins had four degradation peaks (Fig. 8b). The first degradation was detected at around 137°C for Bio-PU-L-EtAc and at 148°C for Bio-PU-L-MeOH, which could be due to the evaporation of residual water and solvent and glass transition of Bio-PU resins. This phenomenon was also detected by the results of DSC (Fig. 6a). At this step, the hard segment of Bio-PU resins changed its phase to a glassy state after the evaporation of residual water and solvent (Gama et al. 2019). The second degradation occurred at around 277°C for Bio-PU-L-EtAc and 282°C for Bio-PU-L-MeOH, which was due to the melting of Bio-PU resins. Further degradation was observed at a temperature around 390°C for both Bio-PU resins. This degradation belonged to the unreacted isocyanate in Bio-PU resins (Gama et al. 2019). The fourth degradation step was detected at around 500°C, which was related to char formation from Bio-PU resins (Gama et al. 2019). The residual mass of Bio-PU resins was quite similar, with around 35% for Bio-PU-L-EtAc and 34% for Bio-PU-L-MeOH.

3.3. Modification of Ramie Fibers

Impregnation of ramie fibers was designed to improve the thermal stability and flame retardancy of ramie as a raw material for functional textiles. Fig. 9 displays the picture of ramie fibers before and after impregnation with Bio-PU resins. The original degummed ramie fibers were white, but the color changed to light brown after the impregnation with Bio-PU-L-EtAc and to dark brown with Bio-PU-L-MeOH. As can be seen in Fig. 1, the Bio-PU resins were brownish,

therefore the color of impregnated ramie fibers changed to light brown and dark brown. The weight gain of ramie fibers after vacuum impregnation was quite similar for both Bio-PU resins at the same impregnation time (**Table 1**). Impregnation of ramie fibers with Bio-PU resins L-MeOH resulted in greater weight gain than those of Bio-PU L-EtAc, regardless of the impregnation time. This is probably due to the fact that more lignin was fractionated by MeOH for the preparation of Bio-PU resins than by EtAc. As can be seen in **Fig. 7**, less free $-NCO$ groups were detected in Bio-PU-L-MeOH than those in Bio-PU-L-EtAc, meaning more Bio-PU resins were produced from L-MeOH. As expected, the weight gain of modified ramie fibers increased with longer impregnation time ([Gindl et al. 2003](#)).



Fig. 9. Photograph of ramie fibers before impregnation (left), after impregnated with Bio-PUL-EtAc (center), and after impregnated with Bio-PU-L-MeOH (right).

Table 1. Weight gain (%) of ramie fibers after impregnation with Bio-PU resins at different times

Bio-PU resins	Time (min)	
	30	60
L-EtAc	25.96 ± 1.08	29.68 ± 0.42
L-MeOH	29.08 ± 0.83	45.47 ± 0.54

As depicted in **Fig. 10**, ramie fibers before the impregnation process had six typical peaks at 3340 cm^{-1} , 2890 cm^{-1} , 1740 cm^{-1} , 1610 cm^{-1} , 1420 cm^{-1} , and 1025 cm^{-1} that belonged to O–H vibration, C–H vibration carbohydrates, C=O vibration of hemicellulose, C–H bending of carbohydrates, C–O of cellulose, respectively ([Kandimalla et al. 2016](#)). After impregnation with Bio-PU resins, the alteration of functional groups of ramie fibers was observed by FTIR. The peak of ramie fibers at 3340 cm^{-1} was shifted to 3330 cm^{-1} due to the reaction of $-OH$ groups of ramie with $-NCO$ groups of Bio-PU (**Fig. 10a**). Two new peaks at 2940 cm^{-1} and 2830 cm^{-1} that assigned to N–H bending and $-CH_2$ stretching of Bio-PU were observed in impregnated ramie fibers with Bio-PU resins. Moreover, peaks at 1740 cm^{-1} and 1610 cm^{-1} of ramie fibers were shifted down to 1710 cm^{-1} and 1600 cm^{-1} , respectively, due to the formation of C=O of urethane of Bio-PU resins (**Fig. 10b**). Therefore, several new peaks were formed in modified ramie fibers such as 1510 cm^{-1} , 1310 cm^{-1} , 1220 cm^{-1} , and 1055 cm^{-1} . Those peaks were corresponding to C–N stretching of primary and secondary amide, C=O vibration from Bio-PU resins, C–O–C of ether linkages, respectively ([Cateto et al. 2008](#)). In addition, peaks at 820 cm^{-1} and 760 cm^{-1} were detected in ramie fibers after impregnation with Bio-PU resins, which originated from $-CH_2$ rocking of isocyanate in Bio-PU resins. This result indicates that free $-NCO$ groups (blue color) of Bio-PU resins reacted with the $-OH$ groups (green color) on the surface of ramie fibers to form urethane linkages (red color), as displayed in **Fig. 11**. It is believed that some of Bio-PU resins were

impregnated into ramie fibers after vacuum impregnation. However, microscopy investigation is needed to determine the depth of impregnation of Bio-PU resins into ramie fibers.

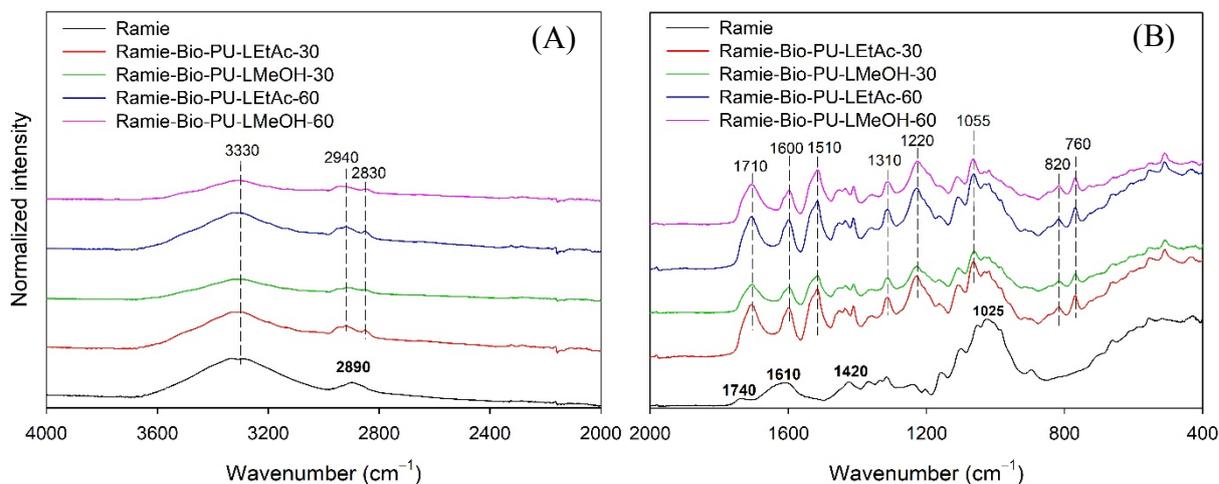


Fig. 10. Functional groups analysis of impregnated ramie fibers. (a) 4000-2000 cm^{-1} , (b) 2000-400 cm^{-1} .

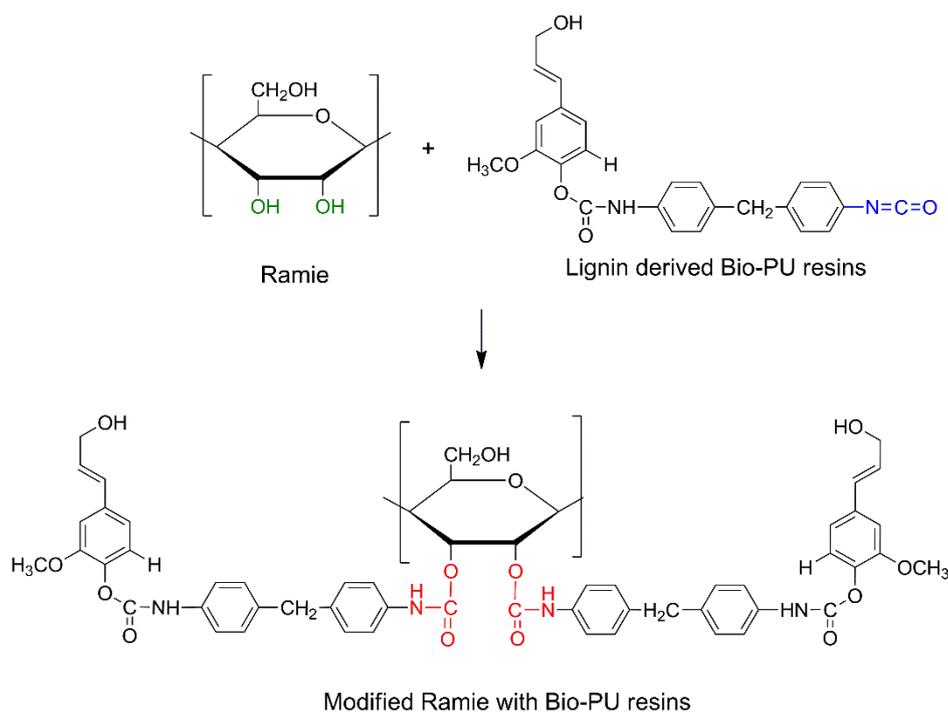


Fig. 11. Scheme reaction of ramie fibers impregnated with Bio-PU resins.

Thermal properties and stability of modified ramie fibers with Bio-PU resins were examined using DSC and TGA-DTG analysis. DSC analysis showed two reactions in ramie fibers, one endothermic reaction and an exothermic reaction (**Fig. 12a**). The endothermic reaction (T_{p1}) occurred at around 46°C owing to moisture evaporation in ramie fibers (Poletto et al. 2015). Modification of ramie fibers with Bio-PU-L-MeOH resulted in similar T_{p1} at around $46\text{--}47^\circ\text{C}$, while impregnation of Bio-PU-L-EtAc into ramie fibers increased the T_{p1} to around 59°C and 88°C . It is known that EtAc has a higher boiling point than the MeOH, around 77.1°C for EtAc and around 64.7°C for MeOH. This could affect the T_{p1} of modified ramie fibers. The exothermic peak (T_{p2}) of ramie fibers took place at around 330°C due to the formation of charring or solid

residue (Shahinur et al. 2020). Impregnation of Bio-PU resins into ramie fibers increased the T_{p2} value to around 345°C until 360°C, where Bio-PU-LEtAc gave higher T_{p2} than that of MeOH.

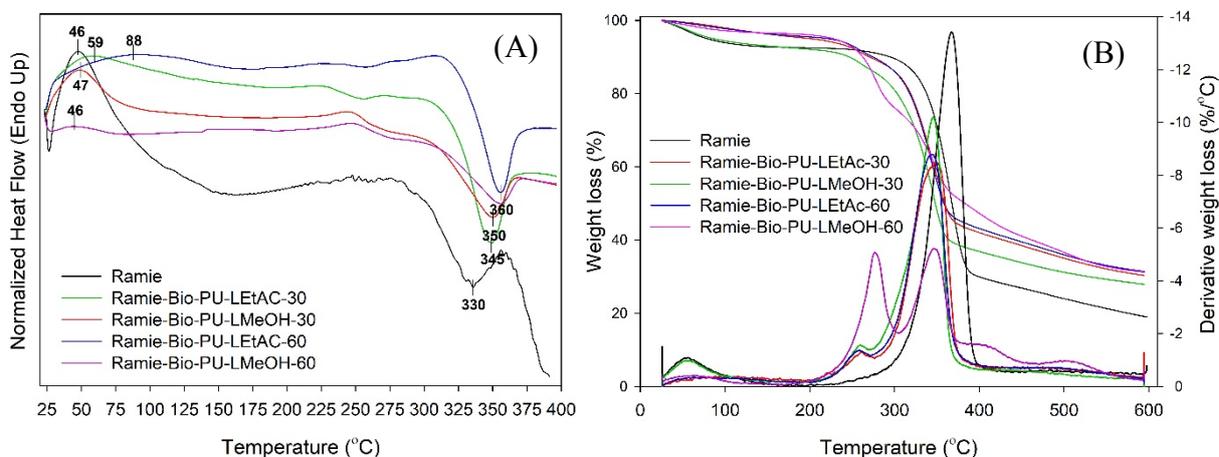


Fig. 12. Thermal properties of ramie fibers before and after impregnation. (a) DSC, (b) TGA-DTG.

TGA-DTG analysis revealed that modified ramie fibers with Bio-PU resins had greater thermal stability than that of original ramie fibers (**Fig. 12b**). The first degradation stage occurred due to water loss and low-molecular-weight components degradation, resulting in nearly 10% of weight loss. Impregnation remarkably increased the first degradation peak temperature from 50°C to around 100°C, resulting in greater thermal stability as indicated by derivative weight loss of around 0.5%/°C. This stage was responsible for the thermal stability, therefore if the fiber is not sufficiently dried, the water contained in the sample can lead to a catastrophic failure when used as composite materials (Poletto et al. 2015). The second degradation stage was attributed to hemicellulose degradation that observed within a narrow range of temperature from 200°C to 300°C, generating around 20-40% of weight loss. The derivative weight loss ranged from 2.0%/°C to 4.5%/°C, where impregnation with Bio-PU-L-MeOH resulted in higher weight loss compared to Bio-PU-L-EtAc. The third stage was related to cellulose degradation at a higher temperature range between 320°C and 400°C, producing around 50-80% of weight loss. The degradation is mainly due to the random cleavage of the glycosidic linkage of cellulose, in which the maximum weight loss generally occurs at 350°C (Poletto et al. 2015). Modification of ramie fibers with Bio-PU resins generated derivative weight loss of around 5.0%/°C to 10%/°C, while the original ramie fibers had around 14%/°C of derivative weight loss. This indicated that the free -NCO groups Bio-PU resins have reacted with the -OH groups of ramie fibers as depicted in **Fig. 11**, thus resulted in greater thermal stability and lower weight loss. The weight residue of ramie fibers was around 18.5%, while the modified ramie fibers had residue of around 28.1-32.5%.

4. Conclusions

This study investigated the possibility of using lignin-derived bio-polyurethane (Bio-PU) resins to modify ramie fibers. Liquid fractions of lignin-ethyl acetate (L-EtAc) and lignin-methanol (L-MeOH) were used as pre-polymer of Bio-PU resins instead of conventional polyol. The Bio-PU resins were successfully prepared by mixing liquid fractions of lignin with 10% of pMDI as indicated by the formation of urethane (R-NH-C=O-R) groups at 1605 cm^{-1} and the

presence of free isocyanate ($-NCO$) groups at $2270-2280\text{ cm}^{-1}$. Modification of ramie fibers with Bio-PU resins using vacuum impregnation method was successfully performed. The formation of urethane ($R-NH-C=O-R$) groups at 1600 cm^{-1} was detected in modified ramie fibers owing to the reaction between free $-NCO$ groups of Bio-PU resins with $-OH$ groups of ramie fibers. Impregnation of ramie fibers with Bio-PU resins remarkably improved the fibers' thermal stability by 10% to 15%, depending on liquid fractions of lignin and the impregnation time. Combination of Bio-PU-L-EtAc and 60 min of impregnation time resulted in higher thermal retardancy ramie fibers compared to the original one. This study showed that lignin derived Bio-PU resins could be successfully used as a fire retardant to improve thermal stability and fire retardancy of ramie fibers.

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